

©2024 College of Veterinary Medicine, Michael Okpara University of Agriculture, Umudike, Nigeria

Original Research

Evaluation of intravenous Acepromazine-Butorphanol-Propofol anaesthesia on Canine serum biochemistry

¹Gapsiso, R.H., ^{2*}Yoksa, D.T. & ¹Mohammed, A.S.A.

¹Department of Animal Health and Production Technology, Federal Polytechnic Mubi PMB 35 Mubi, Adamawa State, ²Department of Veterinary Pathology, Faculty of Veterinary Medicine, University of Maiduguri PMB 1069 Borno State, Nigeria

*Correspondence: dyoksa@gmail.com; +2348065607027

ABSTRACT

In order to increase accuracy and ensure safety during surgical procedures, anaesthesia is a necessary precondition for most diagnostic and surgical procedures in humans and animals. This study was designed to determine the changes in serum biochemical parameters following Acepromazine-Butorphanol-Propofol (ABP-combination) and propofol alone (PRO alone) anaesthesia in dogs. Ten (10) apparently healthy dogs with mean age of 1.59±0.77 years and a mean body weight of 15.5±1.96 kg were obtained from live dog market in Maiduguri, North-eastern Nigeria. The dogs were divided randomly into two groups, ABP-combination and PRO-alone groups. Following premedication with Acepromazine at 0.02mg/kg and Butorphanol at 0.05mg/kg iv., 4mg/kg Propofol was used to induce total intravenous anaesthesia (TIVA) after five minutes of premedication in ABP-combination group, while PRO-alone at 6mg/kg was used to induce anaesthesia without premedication in PRO-alone group. Creatinine (sCr), Alanine aminotransferase (ALT) and Blood urea nitrogen (BUN) were determined using standard laboratory procedures. The ALT, sCr and BUN showed no difference (p>0.05) within the group but were significantly (p<0.05) different between the groups. The levels of Alanine aminotransferase in both ABP and PRO treatments showed significant difference (p<0.05) at 10minutes, 1hour, 6hrs and 24hours. Significant difference (p<0.05) was observed in sCr levels between the two treatments, at 10minutes, 1hour and 6hours respectively. Blood urea nitrogen in both treatments differed (p<0.05) significantly at 6hours and 24hours post anaesthesia. The results of this study showed that ABP-combination following TIVA provides transient non-significant (p>0.05) effect on serum biochemical parameters. Therefore, the combination of ABP can be used during surgical procedures in dog.

Keywords: Acepromazine, anaesthesia, Butorphanol, dogs, Propofol, serum biochemistry.

INTRODUCTION

Controlled, reversible unconsciousness, analgesia, and muscle relaxation are the hallmarks of anaesthesia (Thurmon & Short, 2007). Improving surgical accuracy is an essential precondition for the majority of medical and diagnostic procedures in humans and animals (De Moor *et al*, 2010, Elks, 2014,).

Over time, intravenous anaesthesia has gained popularity and acceptance in veterinary practice as a more acceptable method of achieving ideal surgical anaesthesia in comparison with inhalational anaesthesia, which necessitates the use of multiple facilities for the delivery of volatile anaesthetic agents and the potential hazard to the patient during administration (Umar et al., 2006; Yamashita et al., 2007; Umar 2007; al., et al., Dantino et 2022). Combining analgesic, muscle relaxant, and sedative drugs -

properties that are rarely offered by a single anaesthetic drug- is the usual goal of the anaesthetic agent combining technique in order to achieve balanced anaesthesia (Waelbers et al., 2009; Morton and Hall 2012; Umar et al., 2007). During general anaesthesia, the combination of anaesthetic drugs is typically administered in a dosage that maximizes the desired or major effects while minimizing the negative effects. This allows the beneficial effects of each anaesthetic agent to be utilized to achieve an appropriate depth of surgical anaesthesia while causing predictable and minimal effects on the organs involved in drug metabolism and excretion (Thurmon & Short, 2007; Gapsiso et al., 2023).

To attain balanced anaesthesia, total intravenous anaesthesia (TIVA) should be used in conjunction with other drug

combinations and techniques that are appropriate for the ever-evolving demands of advanced diagnostic and therapeutic modalities (Waelbers et al. 2009; Bajwa et al. 2010). Propofol (2,6-diisopropylphenol) is an intravenous phenolic compound that is non-barbiturate. It induces anaesthesia smoothly and quickly, resulting in a brief period of unconsciousness followed by а swift and uneventful recovery (Hall et al., 2001; Umar et al., 2006). One phenothiazine tranquillizer that is frequently used for premedication and mild sedation is acepromazine, a phenothiazine that does not produce analgesia but facilitates the induction of anaesthesia at clinically prescribed doses (Hall et al., (2001). On the other hand, butorphanol is a synthetic agonist-antagonist opioid analgesic of the morphine type that is frequently used as a supplement for balanced general anaesthesia and for the treatment of moderate-to-severe pain (Elks, 2014).

In order to provide balanced anaesthesia in dogs and to evaluate acepromazine, butorphanol and propofol's effects metabolism and excretion organs, on drug in addition to paucity of information regarding ABP drug combinations and Propofol-alone (PRO) for anaesthesia in dogs in the study area, necessitated the choice of the drugs for TIVA. The purpose of the study was to assess how the ABP combination affected the serum biochemical markers creatinine, blood urea nitrogen, and alanine aminotransferase in dogs.

METHODOLOGY

EXPERIMENTAL ANIMALS AND MANAGEMENT

All procedures performed in this experimental study were in accordance to the guidelines of the animal care and use committee of the University of Maiduguri (ethical approval number: FVM/UM/AUEC/19/002). Ten (10) apparently healthy dogs comprising six males and four female of good clinical condition and having mean body weights of 15.5 \pm 1.96 kg and age 1.59 ± 0.77 years old were used. The dogs were kept at the kennels and fed corn food and rice once daily and clean water was provided ad libitum. Following a two-week acclimatization, period of they were randomized into two treatment groups, the Acepromazine-Butorphanol-Propofol (ABP) combination group and the Propofol (PRO) alone group.

Three drugs Acepromazine, (Neurotranq[®] 10mg/ml Virbac, RSA Pty. Ltd. South Africa), Butorphanol (Dolorex[®] 10mg/ml Intervet SA Pty. Ltd. South Africa) and Propofol[®] 1% Frensenius Kabi SA Pty. Ltd. South Africa) were used in two treatment groups: ABP-combination and PRO-alone groups for the anaesthesia to which the dogs were assigned.

Animals assigned to ABP group were premedicated with acepromazine (0.02mg/kg iv) and butorphanol (0.05mg/kg iv). Using insulin syringes, the calculated doses of the

two medications for premedication were taken from separate vials and combined into a single 5 ml syringe. Water for injection was then added to the mixture to make up to 1 ml before the mixture was administered intravenously (iv) through a $22G \times 0.9$ DEN-V i.v. cannula (Polybond India Pvt Ltd. Maharashtra India) inserted into the cephalic vein, 27.0 ± 1.0 seconds after, the induction of anaesthesia with propofol (4 mg/kg) iv followed. In both groups, 2ml of blood was collected before premedication and during the anaesthesia at 10minutes interval and at 1hr, 6hrs and 24hrs post-anaesthesia, thereafter sera samples were harvested. The dogs assigned to propofol-alone group were not premedicated but anaesthesia was induced with 6mg/kg of propofol intravenously through cephalic vein.

MEASUREMENT OF SERUM BIOCHEMICAL PARAMETERS

Alanine aminotransferase, creatinine and blood urea nitrogen were measured in the sera samples collected. The blood samples were collected before the commencement of the treatments for baseline values and later collected at 10 minutes intervals during anaesthesia and then 1 hour, 6 hours and 24 hours after the anaesthesia. The blood samples were gently dispensed into well labelled plain vacutainer tubes and allowed to clot for 2 hours at room temperature thereafter centrifuged 1200xG for 5 minutes. The sera were thereafter harvested into micro-vial (1ml) tubes and stored at -20°C before analysis. Alanine-aminotransferase (ALT) was determined using the method of Schmidt and Schmidt (1963) and the concentration of ALT in the serum was calculated from a standard table (Randox Laboratory Ltd. U.K.). Serum Creatinine (sCr) was determined using Randox Laboratory kits and procedures (Natelson et al., 1951). The Blood Urea Nitrogen (BUN) was determined by diacetyl method of Natelson et al., (1951) using Randox Laboratory Kits and procedures.

STATISTICAL ANALYSIS

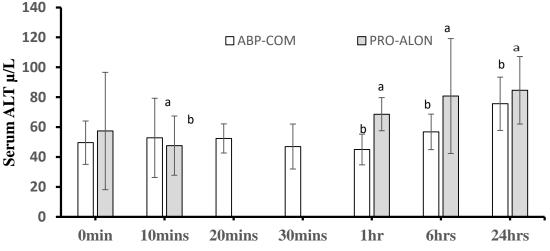
The data were presented as mean \pm SD. One Way Repeated Measure Analysis of variance (ANOVA) was used to analyse data within group while student T-Test was used to analyse data between the groups. Analyses were considered significant at p < 0.05.

RESULTS

The duration of anaesthesia for ABP-combination was 30.0 ± 5.8 minutes while that of PRO-alone was 13.6 ± 4.8 minutes, therefore, no values were obtained for PRO at 20 minutes and 30 minutes due to shorter duration of anaesthesia.

The baseline value for ABP-combination was $49.6\pm14.5\mu/L$. There was a non-significant (p > 0.05) decrease in ALT level from 10 minutes to 1 hour following the treatment in ABPcombination. The level rose insignificantly (p > 0.05) at 6 hours and at 24 hours above the baseline (Fig 1). The changes in the ALT levels did not differ significantly (p > 0.05) from the baseline. The baseline ALT value for PROalone was 57.4 \pm 39.2 µ/L and there were no significant (p > 0.05) changes in ALT values during the sampling time from the baseline. Comparing the results on the effects of the two treatments on ALT showed significant (p < 0.05) difference between ABP-combination and PRO-alone at 10 minutes during anaesthesia, 1 hour, 6hours and 24 hours post anaesthesia. ABP-combination showed increase from its 24 hours post anaesthesia which did not differ significantly (p > 0.05) between them (Figure II.).

The baseline value of Blood urea nitrogen BUN for ABPcombination was $3.9 \pm 1.3 \ \mu mol/L$). There was nonsignificant (p > 0.05) decrease in BUN levels at 10 minutes, 20 minutes and 30 minutes followed by non-significant (p > 0.05) rise in the BUN level at 1 hour, 6 hour and 24 hours post anaesthesia. The fluctuation in the BUN level did not differ significantly (p > 0.05) from the baseline (Fig 3). The baseline value of BUN for PRO-alone treatment was 4.4 ± 1.6 µmol/L. The BUN level dropped non-significantly (p >



Time interval

Fig. 1. Effect of ABP-Combination and PRO-Alone on Alanine aminotransferase (ALT) in dogs. Values with different alphabet for a Time interval are significantly different (p<0.05)

baseline while PRO-alone showed decrease from its baseline (Figure I).

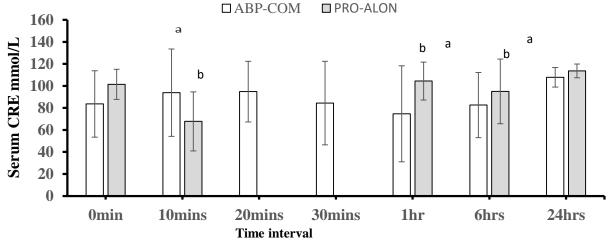
The serum creatinine (sCr) baseline value obtained from the study for ABP-combination was 83.6 ± 30.1 mmol/L. There was slight increase in sCr levels at 10 minute and 20 minutes respectively from baseline. There was decrease in sCr level observed at 30 minutes, 1 hour and 6 hours respectively. The sCr level increased to a peak in 24 hours, which was higher than the baseline value. The changes in sCr levels recorded at various sampling times within the ABP-combination group did not differ significantly (p > 0.05) from the

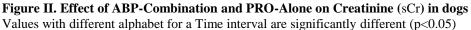
baseline. The baseline value of sCr for PRO-alone was 101.4 \pm 13.7 mmol/L. There was decrease sCr level at 10 minutes, and the sCr level rose at 1 hour, followed by dropped at 6 hours and later increased at 24 hours. No values for PRO-alone at 20 minutes and 30 minutes during anaesthesia due to shorter duration of anaesthesia with PRO-alone. All the sCr values for PRO-alone recorded did not differ significantly (p>0.05) from the baseline (Fig 2). The two treatments recorded increased sCr values above their baseline values at

0.05) at 10 minutes and 1 hour, rose at 6 hours'and dropped again at 24 hours. Comparing the effects of ABP-combination with PRO-alone on BUN showed significant (p < 0.05) difference in BUN values at 6 hours and 24 hours (Fig 3.).

DISCUSSION

The values for ALT within the groups did not differ significantly (p > 0.05) but the effect of the two groups when compared, vary significantly (p < 0.05) with time. This finding agreed with the report of Anandmay, *et al.*, (2012), who reported that a variable and consistent non-significant (p > 0.05) increase in ALT, which were within the physiological limits, could be noticed at different intervals in dogs.. The findings of this study varied with the findings of Dewangan *et al.*, (2016) who reported a significant (p < 0.01) variation in serum ALT values between groups following xylazine and propofol anaesthesia in dogs. Shabir *et al.* (2014) also reported a significant (p < 0.05) difference





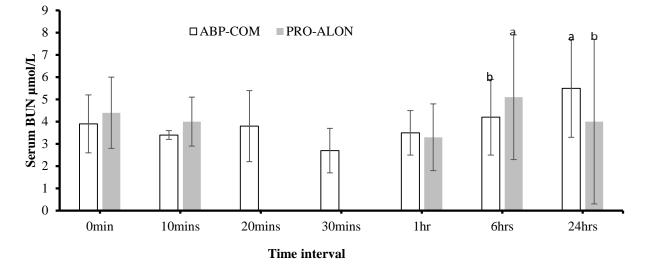


Figure III. Effect of ABP-Combination and PRO-Alone on Blood Urea Nitrogen (BUN) in dogs. Values with different alphabet for a Time interval are significantly different (p<0.05)

in ALT levels following propofol bolus induction in humans. The transient non-significant increase in ALT level recorded in this study could be attributed to the hepato-tolerance activities of acepromazine present in the combination which help prevent any possible hepatic injury or metabolism of the drug by the liver (Cattai *et al.*, 20018). Because of the antisympathetic and antiarrhythmic properties and dopamine inhibition activity of acepromazine, the drug help stabilize drug metabolic activities of liver by inducing vagally-bradicardia followed by systemic vasodilatation thereby slows down excessive flow of blood to the liver, which in turn reduces hepato-toxicity (Turi & Williams, 2011). The findings of this study differ with the findings of Anandmay *et al.*, (2012), who reported a significant (p < 0.05) increase in ALT level at 1 and 2 hours post induction of propofol

which later return to the baseline within 24 hours after the induction. In the present study, both treatments recorded non-significant (p > 0.05) increase in ALT values above baseline values 24 hours post anaesthesia. The transient increase in ALT values in ABP-combination group could be due to hepatic metabolism of acepromazine and butorphanol which returned back to normal physiological level indicating no undesirable effect (Anandmay *et al.*, (2016).

The non-significant (p > 0.05) variations in sCr levels recorded with ABP-combination in this study could be attributed to the increased level of antidiuretic hormone (ADH) along with decreased glomerular filtration activity during anaesthesia procedure in dogs (Suresha *et al.*, 2012). ABP-combination recorded initial non-significant (p > 0.05) increase in creatinine while PRO-alone recorded nonsignificant (p > 0.05) decrease in creatinine value from baseline after induction of anaesthesia. The variation in sCr values recorded between the two treatments could be attributed to the metabolism, bioavailability and half-life of the acepromazine (6 to 8 hours) and butorphanol (3 to 4 hours) in dogs which is longer than propofol (40minutes) as reported by Morton, (1998); Hammond & Nickelson (2008); Maddison *et al.*, (2008); Hall *et al* (2001).

The non-significant (p > 0.05) variation in BUN values recorded post anaesthesia between the two treatments could be due to the longer half-life of acepromazine (6 to 8 hours) and butorphanol (3 to 4 hours) which is longer than propofol (40minutes) alone in dogs (Morton, 1998; Hammond and Nickelson 2008; Maddison et al., 2008; Hall et al 2001). The non-significant (p > 0.05) changes in BUN values recorded during anaesthesia in this study shows non-nepro-toxicity of the anaesthestic agent which are usually observed following drug metabolism and the half-life of the anaesthetics (acepromazine has 6 to 8 hours, butorphanol has 3 to 4 hours while Propofol has about 40minutes in dogs) which vary (Bougherara & Bouaziz 2014). Acepromazine causes vasodilatation which increases blood flow to the kidneys while butorphanol often increases left ventricular enddiastolic pressure and an increased systemic arterial pressure brings about increase blood flow to the kidneys thereby reducing any deleterious injury to the kidneys (Hall et al., 2001). This could be the reason for the non-significant (p >0.05) changes in the BUN level recorded in this study.

CONCLUSION

Both ABP combination and PRO-alone recorded transient non-significant effects on the liver and kidney during and after anaesthesia which indicates their suitability for use in dogs. The non-significant fluctuation in the ALT, sCr and BUN within the groups during after anaesthesia, shows that the ABP combination and PRO-alone doses used did not cause any deleterious effect or changes in the serum biochemical parameters. Nevertheless, the significant changes recorded between the groups were as a result of differences in doses of the propofol in the two groups and the inclusion of acepromazine and butorphanol in ABP combination. At the doses used in this study, ABP combination and propofol-alone are safe and can be used in dogs. Further clinical and surgical trials are recommended to determine quality of anaesthesia in other species of animals.

ACKNOWLEDGEMENT

The authors acknowledge the Department of Animal Health and Production Technology, Federal Polytechnic Mubi, Department of Veterinary Pathology, University of Maiduguri and Department of Veterinary Surgery and Radiology, University of Maiduguri for providing facilities for the research.

CONFLICT OF INTEREST

The authors declare that they have no conflict of interest

REFERENCES

- Anandmay, A. K., Dass, L. L. and Sharma, A. K. (2012). Clinico-anaesthetic changes following administration of propofol alone and in combnation of buprenorphine in dogs. *Indian Veterinary Journal*, 89(10), 77-79.
- Anandmay, A. K., Dass, L. L., Sharma, A. K. and Gupta, M. K. (2016). Haemato-biochemical changes following administration of propofol in combination with buprenorphine in atropinized dogs. *Journal of Animal Research*, 6(3), 531-36.
- Bajwa, S. J., Bajwa, S. K. and Kaue, J. (2010). Combination of two drugs combination in total intravenous anaesthetics: propofol-ketamine and propofolfentanyl. *Saudi Arabian Journal of Anaesthesia*, 4(2), 72-79.
- Bougherara, H. and Bouaziz, O. (2014). Effects of the anaesthetic/ tranquilizer treatments (ketamine, ketamine+acepromazine, zoletil) on selected plasma biochemical parameters in laboratory rats. *Central European Journal of Experimental Biology*, 3(2), 1-5.
- Cattai, A., Rabozzi, R., Ferasin, H., Isola, M. and Franci, P. (2018). Haemodynamic changes during propofol induction in dogs. new findings and approach of monitoring. *BMC Veterinary Rsearch*, 64(5), 590-598.
- Dantino, S.C., Kleine, S.A., Smith, C.K., Smith, S.M., Zhu X. and Sededighi, R. (2022). ffects of intravenous acepromazine and butorphanol on propofol dosage for induction of anaesthesia in Beagle dogs. *Veterinary Anaesthesia and Analgesia*. 49(4), 354-363. https://doi.org/10.1016/j.vaa.2022.03.002
- De Moor. A., Verschooten, F., Desmet, P. and Steenhaunt, M. (2010). Intrathoracic cardiac resucitation in the horses. *Equine Veterinary Journal*, 4(1), 31-33.
- Dewangan, R., Tiwari, S. K. and Sharda, R. (2016). Haemobiochemical response to xylazine -propofol anaesthesia and its reversal by yohimbine and atipamezole in dogs. *International Journal of Science and Nature*, 7(3), 647-650.
- Elks, J. (2014). *The dictionary of drugs: chemical data, structures and bibliographies.* Springer Science Business Media. Dordrecht: 114.
- Fleckmell, P., Jennifer, L. S., Melissa, C. D. and Roberth R. M. (2015). Pre-anaesthesia, anaesthesia, analgesia and euthanasia. In *Laboratory Animal Medicine* (3rd ed. 1135-1200).
- Gapsiso, R.H., Umar, M.A and Mohammed, A. (2023). Evaluation of anaesthetic indices and physiological variables following total intravenous anaesthesia with Acepromazing-Butorphanol-Propofol combination in dogs. Sahel Journal of Veterinary Sciences. 20(1)
- Hall, L. W. and Clarke, K. W. (1991). General Pharmacology of Intravenous Anaesthetics Agents. In L. W. Hall, *Veterinary Anaesthesia* (9th ed.,). London: Bailliere Tindall. 522-525.

- Hall, L. W., Clarke, K. W. and Trim, C. M. (2001). Veterinary Anaesthesia (10th ed.). Edingburg, UK.: WB Sauders, 80-97.
- Hammond, R. and Nicholson, A. (2008). Small Animal Clinical Pharmacology (2nd ed.). (J. E. Maddison, Ed.) Philadelphia.: Saunders Limited. doi:https://doi.org/10.1016/B978-0-7020-2858-8.x5001-5
- IBM Statistical package for social sciences, 2011 (Version 20.0).
- Morton. I. K. and Hall, J. M. (2012). Concise Dictionary of Pharmacological Agents: Properties and Synonyms. Neitherlands: Springer Science and Business Media. 101.
- Natelson S., Scott, M.L. and Beffa, C. (1951) A rapid method for the estimation of urea in biologic fluids. *American Journal of Clinical Pathology*. 21(3): 275-281.
- Schmidt, E. and Schmidt, F. W. (1963). Glutamic-pyruvic transaminase determination. *Enzymology, Biological and Clincals.* 3, 1. 245-256
- Shabir, N., Iqbal, R. and Abbas, M. N. (2014). Effects of propofol on haemodynamics and blood profile of humans. *World Journal of Medical Science*, 10(1), 74-77. doi:10:5829/idosi.wims.10.1.828
- Suresha, L., Ranganath, B. N., Vasanth, , M. S. and Ranganath, L. (2012). Haemo-biochemical studies on triflupromazine HCL and diazepam premedication for propofol anaesthesia in dogs. *Vet. World.*, 5(11), 672-675.
- Thurmon, J. C. and Short, C. E. (2007). History and overview of veterinary anaesthesia. In G. K. C., *Lumbs and Jones Veterinary Anaesthesia and Analgesia* (4th ed.,). Oxford: Blackwell Publishing Ltd. 3-6.

- Turi, K. A. and William W.M. (2011). Pain assessment and management. Small Animal Paediatrics. The first 12 months of life. Michael E. Peterson and Michelle A. Kutzler ISBN 978-1-4160-4889-3 Elsevier Inc. Doi: https://doi.org/10.1016/C2009-0-37825-3
- Umar, M. A., Yamashita, K., Kushiro, T. and Muir, W. W. (2006). Evaluation of total intravenous anaesthesia with propofol or ketamine-meditomidine-propofol combination in horses. *Journal of American Veterinary Medical Association.*, 228(8), 1221-1227.
- Umar, M. A., Yamashita, K., Kushiro, T. and Muir, W. W. (2007). Evaluation of cardiovascular effects of total intravenous anaesthesia with propofol or a combimation of ketamine-meditomidine-propofol in horses. *Asian Journal of Veterinary Research*, 68(2), 121-126.
- UN Morld Urbanization Prospects 2017
- Waelbers, Y., Vermoere, P, and Polis, I. (2009). Total intravenous anaesthesia in dogs:. *Diergenees Kundig Tijdschrift Journal*, 78, 160-169.
- Yamashita, K. Wijayathilaka, T. P., Kushiro, T., Umar, M. A., Taguchi and Muir, W. W. (2007). Anaesthetic and cardiopulmonary effects of total intravenous anaethesia using midazolam, ketamine and meditomidine drug combination in horses. *Journal of Veterinary Medical science*, 69(1), 7-13.